simplifying research

qPCRBIO Probe 1-Step Go Lo-ROX www.pcrbio.com

# Product description:

qPCRBIO Probe 1-Step Go is a universal probe kit designed for fast, highly specific and ultra-sensitive RT-qPCR. The latest developments in reverse transcriptase technology and buffer chemistry are used to give efficient cDNA synthesis and real-time PCR in a single tube.

The kit is engineered for use on a wide range of probe technologies such as TaqMan®, Scorpions® and molecular beacon probes. It can be used to quantify any RNA template including mRNA, total RNA and viral RNA sequences. qPCRBIO Probe 1-Step Go is designed to give rapid and accurate results over a broad range of template concentrations and is ideally suited to the detection of RNA viruses including SARS-CoV-2.

The kit includes a thermostable and extremely active modified MMLV reverse transcriptase (RTase Go) and advanced RNase inhibitor that prevents degradation of RNA by contaminating RNase. Antibody-mediated hot start technology prevents the formation of primer dimers and non-specific products giving highly specific and ultra-sensitive real-time RT-PCR with unrivalled efficiency in multiplex.

High-throughput screening has resulted in a buffer system that allows efficient amplification from GC-rich and AT-rich templates, under fast and standard cycling conditions.

witec ag
Industriestrasse 12   6210 Sursee T 041 250 53 57   mail@witec.ch

Pack size	2x qPCRBIO Probe 1-Step Go Lo-ROX	20x RTase Go (with RNase inhibitor)
100 reactions	1 x 1mL	1 x 100μL
300 reactions	3 x lmL	3 x 100µL
500 reactions	1 x 5mL	1 x 500μL
1200 reactions	12 x 1mL	12 x 100μL
5000 reactions	1 x 50mL	1 x 5mL
50000 reactions	1 x 500mL	1 x 50mL

## Shipping and storage

On arrival the kit should be stored between -30°C and -15°C. Avoid prolonged exposure to light. If stored correctly the kit will retain full activity for 12 months. The kit can go through 30 freeze/thaw cycles with no loss of activity.

# Limitations of product use

The product may be used for in vitro research purposes only.

# Technical support

Help is available on our website at https://pcrbio.com/resources/ including answers to frequently asked technical questions. For technical support and troubleshooting please email technical@pcrbio.com with the following information:

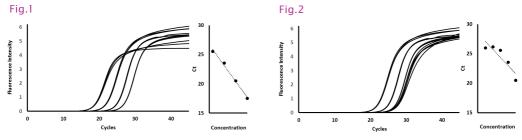
- Amplicon size
- Reaction setup
- Cycling conditions
- Screen grabs of amplification traces and melting profile

### Important considerations

Instrument compatibility: Different real-time PCR instruments require different levels of ROX passive reference. Generally, modern instruments do not require passive reference but include the option to use it for normalisation. Please check our qPCRBIO Selection Table to determine which ROX concentration your instrument requires (https://pcrbio.com/resources/qpcr-selection-tool/).

Primer design: For efficient amplification under fast cycling conditions we recommend amplicon lengths between 80bp and 200bp. With all manufacturers, the shorter the amplicon length, the faster the reaction can be cycled. Amplicon lengths should not exceed 400bp. Primers should have a predicted melting temperature of around 60°C, using default Primer 3 settings (http://bioinfo.ut.ee/primer3/). For TaqMan® probes, choose a probe close to the 5' primer and avoid terminal guanosine residues.

Template concentration: As target copy number will vary, it is important to select the correct template concentration to correctly quantify the target sequence. A good concentration will display clear separation between amplification curves (Fig.1). At lower template concentrations, the amplification curves will begin to group together and Ct values will not fit the standard curve (Fig.2).



#### Reaction setup

- 1. Before starting, briefly vortex 2x qPCRBIO Probe 1-Step Go Mix.
- 2. Prepare a master mix based on the following table. We also recommend setting up a no-RTase control:

Reagent	20µL reaction Final co		Notes
2x qPCRBIO Probe 1-Step Go Mix	10μL	1x	
Forward primer (10µM)	0.8µL	400nM	Con about fav autimal unimas design
Reverse primer (10µM)	0.8μL 400nM		See above for optimal primer design
Probe (10µM)	0.4µL	200nM	
20x RTase Go	0.2µL	0.2x	0.2µL for sensitive SARS-CoV-2 detection. Alternatively, titrate down to 0.05uL.
Template RNA	Viral RNA: 10 to 1x10 <sup>s</sup> copies Total RNA: 1pg to 1µg mRNA: >0.01pg	Variable	Addition of sample as 2 to 5µL volumes will improve assay precision. 5µL of swab extract is recommended for SARS-CoV-2 diagnostic assays
PCR grade dH <sub>2</sub> O	Up to 20µL final volume	• • • • • • • • • • • • • • • • • • • •	

3. Program the instrument using the following conditions, acquiring data on the appropriate channel:

Cycles	Temperature	Time	Notes
1	45°C to 55°C	10min	Reverse transcription: 45°C is recommended for most applications. 55°C should be used only when amplicon contains regions of high secondary structure
1	95°C	2min	Polymerase activation
40	95°C 60°C to 65°C	5 seconds 20-30 seconds	Denaturation Anneal/Extension: do not exceed 30 seconds, do not use temperatures below 60°C
Melt analysis	s Refer to instrument instructions		Optional melt profile analysis, available for hybridisation probes only